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Publication number:

0 381 490
A2

EUROPEAN PATENT APPLICATION

Application number: 90301044.5

Int. Cl.⁵ **A61K 48/00**

Date of filing: 01.02.90

Priority: 02.02.89 US 305856

Date of publication of application:
08.08.90 Bulletin 90/32

Designated Contracting States:
DE ES FR GB IT NL

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Gene therapy using stromal cells.

Production and secretion into the bloodstream of a human patient of a biologically active enzyme for which the human patient suffers a deficiency is achieved by introducing into the human patient donor bone marrow stromal cells which have been transfected with a gene encoding the enzyme, so that the introduced cells can adhere to a bone cavity surface of the patient and produce and secrete the active enzyme.

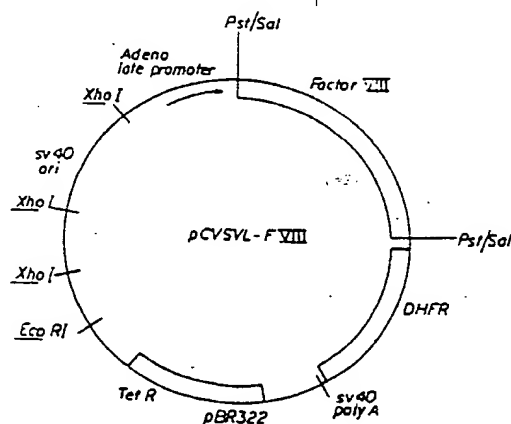


FIG.3

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GENE THERAPY USING STROMAL CELLS

This invention relates to the use of recombinant techniques to correct genetic deficiencies, and more particularly to the use of donor bone marrow stromal cells in therapy.

Hemophilia A, a bleeding disorder caused by a deficiency or abnormality of a particular clotting protein, Factor VIII-C, occurs in about 10-20 males in every 100,000. Afflicted individuals suffer episodes of uncontrolled bleeding and are treated currently with concentrates rich in Factor VIII-C derived from human plasma. Recombinant DNA technology has been proposed as useful for providing human Factor VIII-C purified from cultured cells as an alternative treatment for hemophiliacs (Toole et al., 1984, Nature 312:342).

Louis et al. (1988, Proc. Nat. Aca. Sci. 85:3150) implanted collagen-embedded mouse primary skin fibroblasts infected with a recombinant retrovirus containing human factor IX cDNA in the epidermis of mice, and detected human factor IX for 10-12 days in the sera.

Garver et al. (1987, Science 237:762) transplanted murine fibroblasts containing recombinant DNA encoding human α_1 -antitrypsin into the peritoneal cavity of nude mice, and detected human α_1 -antitrypsin in the sera and the epithelial surface of the lungs. Garver et al. observe that "the level of gene expression in target cells such as fibroblasts has generally been better than with primate bone marrow stem cells".

In general, the invention features a method of causing production and secretion into the bloodstream of a human patient of a biologically active enzyme for which the human patient suffers a deficiency; the method involves introducing into the human patient donor bone marrow stromal cells which have been transfected with a gene encoding the enzyme, so that the introduced cells can adhere to a bone cavity surface of the patient and produce and secrete the active enzyme.

In preferred embodiments, the transfected gene encodes a lymphokine, a growth factor, or a hematopoietic factor, or another deficient blood-borne protein, such as a coagulation factor, e.g., human Factor VIII-C or anti-Thrombin III. The donor stromal cells are preferably autologous cells, i.e., cells that are genetically identical to the recipient cells, so that immunologic rejection of the engrafted donor cells is minimized; if the donor cells are not autologous, they are preferably similar enough genetically to the recipient to avoid an unwanted immune response; i.e. heterologous cells that are histocompatible, may be used. The use of autologous or histocompatible cells obviates general immune suppression of the recipient.

Preferably, the interaction between the donor stromal cells and the recipient bone is altered to facilitate adherence and stable engraftment by irradiation (preferably X-irradiation) of the recipient bone at the site into which the donor stromal cells are to migrate once they are introduced into the animal. The X-irradiation dosage used is high enough to kill hematopoietic cells, so that the majority of rapidly dividing cells are cleared from the site, and to damage stromal cells at the site, but low enough to avoid death of those stromal cells. The site is preferably a long bone, such as a femur, tibia, or rib.

The invention can be used to treat Factor VIII-deficient hemophilia without eliciting an unwanted immune response to the replenished Factor VIII, because the method of transfer of the Factor VIII gene into the recipient is by autologous transplantation of cells capable of expressing the Factor VIII gene. This is in contrast to other methods for treating hemophilia, which have serious disadvantages. For example, human plasma is both an inefficient and expensive source of Factor VIII, and contains only 100-200 ng/ml of the protein (Fay et al., 1982, Proc. Nat. Aca. Sci. 79:7200). The available therapy, although reasonably effective is very costly and is associated with a high risk of infection. The plasma-derived products currently available are highly impure (<1% Factor VIII) and are commonly produced from pooled plasma lots derived from thousands of donors. These products are associated with a variety of serious complications caused by protein precipitates and are frequently contaminated with adventitious agents such as hepatitis virus and human immunodeficiency virus, the causative agent in acquired immune deficiency syndrome (AIDS). Recombinant Factor VIII, produced by eukaryotic cells transfected with the human Factor VIII-C gene (Wood et al., 1984, Nature 312:330; Toole et al., *supra*), has not been purified to homogeneity, and, because it may differ from natural human Factor VIII-C in, e.g., glycosylation pattern, may be immunogenic. The invention also advantageously uses stromal cells, which are long-lived and not rapidly dividing, and are therefore capable, after engraftment, of expressing and secreting the desired enzyme for a prolonged period of time.

Other features and advantages of the invention will be apparent from the following description of the preferred embodiments thereof.

The drawings will first briefly be described.

Fig. 1 is an illustration of explanted adherent cells from a transplanted donor mouse at 1 month after engraftment (upper), or from a non-transplanted-irradiated mouse (lower).

Fig. 2 is a diagram of hematopoietic progenitor cell growth from LTBMCS established from S1 S1^d

mice.

Fig. 3 is a diagrammatic representation of a retroviral vector containing the human Factor VIII.

As is discussed above, a human patient suffering from a genetic deficiency disease caused by the absence of a normally functioning enzyme, e.g., hemophilia caused by deficiency of normal Factor VIII C, can be treated, by transfecting autologous human stromal cells with a vector containing the human Factor VIII C gene, and then engrafting these transfected stromal cells into a suitably prepared human patient, e.g., a patient who has had irradiation treatment of a target site in a long bone.

Before describing in detail the procedure for treating human hemophilia, there will be described, for purposes of enabling some of the techniques employed, an experiment in which neomycin-resistant stromal cells were engrafted into mice; this work is described in the prior art publication Anklesaria et al., (1987) Proc. Nat. Acad. Sci. USA 84, 7681. There will then be described the transfection of murine stromal cells with a gene encoding transforming growth factor- α (TGF- α), followed by the engraftment of those stromal cells into mice; the engrafted cells were found to express and secrete the TGF- α encoded by the introduced gene. Finally, there will be described the procedures by which human hemophilia will be treated, beginning with the initial engraftment into mice of Factor VIII gene-containing stromal cells, followed by the engraftment of Factor VIII gene-containing autologous human cells into human patients.

Example 1

The Anklesaria et al. Experiments

Long-term Cell Cultures

B6D2F1 (C57BL/6J x DBA/2J) mice (Jackson Laboratories, Bar Harbor, Maine) were used to establish long term bone marrow cultures (LTBMC's). The cellular contents of a femur and tibia were flushed into a 25-cm² Corning flask in Fisher's medium (Gibco) supplemented with 25% horse serum (Hazelton) and 10⁻⁶ M hydrocortisone sodium succinate. Fetal calf serum was substituted for horse serum after two weeks. This method has been shown to increase culture longevity (Sakakeeny et al., 1982, J. Nat. Cancer Inst. 68:305). LTBMCs were infected after seven days in culture, and for three consecutive weeks, with viral supernatants containing virus packaged as pure defective virus and 2 μ g/ml polybrene (Sigma).

Establishment of a Stromal Cell Line

On day 61 after establishment of LTBMCs, adherent cells from representative LTBMC flasks were removed by treatment with 0.25% trypsin (GIBCO) and replated at either 3 x 10⁶ or 1 x 10⁶ cells/25-cm² flask (Falcon) in 8.0 ml Fisher's medium (GIBCO) supplemented with 25% heat-inactivated fetal calf serum (FCS) (Hazelton) and 10⁻⁶ M hydrocortisone sodium succinate. Cultures were maintained at 33°C, 7% CO₂, and passaged weekly at the same densities. At passage 8, each of the developing cell lines was plated at limiting dilution and cloned by using penicyclinders to separate single-cell derived colonies. Each of the subsequent cell lines was further subcloned twice by limiting dilution. The cell lines were contact inhibited when they reached confluency. All stromal cell lines tested were found to be reverse transcriptase negative, as well as mycoplasma free. Reverse transcriptase activity was measured according to Greenberger, 1984, In: Golde (ed.) Meth. Hemotol. 11, N.Y.: Churchill Livingston, p.202, using NIH/3T3 cells as negative controls.

Two clonal murine bone marrow stromal cell lines, GBL/6 and GBL/6 neo', were developed from continuous bone marrow cultures of B6.cast-GPIA mice (Glu6PI- α , D-glucose-6-phosphate ketol-isomerase; EC 6.3.1.9) mice (Greenberger, 1978, Nature 275:752). The GBL/6 cell line was chosen for these studies because of its endothelial-like characteristics and in vitro support capacity for multipotential hematopoietic progenitor cells.

The stable clonal stromal cell line GBL/6 expresses the α isoenzyme of Glu6PI. The cell line was characterized by staining for alkaline phosphatase, acid phosphatase, peroxidase, α -naphthyl esterase, and lysozyme, as described in Pearse, 1986, Histochem., I, Churchill, London, 3rd ed. Antisera to extracellular

matrix proteins laminin, fibronectin (Collaborative Research, Waltham, MA), collagen types I and IV, and the alloenzyme marker Glu6PI- α (Charles et al., 1980, Mol. Cell. Biochem. 29:11) were used to identify each protein in the GBL/6 cell line using each specific antiserum and the immunoperoxidase technique of Hadji et al., 1983, Lab. Med. 14:767. GBL/6 cells were fibronectin +, laminin +, and collagen-type IV +, collagen type I +. For immunohistological studies, proximal tibiae were collected, split longitudinally, and fixed for 18 hr in 2% paraformaldehyde/0.1 M cacodylate buffer, pH 7.4. Bones were decalcified in 0.3 M EDTA/0.1 M cacodylate buffer, pH 7.4 for 4 days. Paraffin-embedded bones were sectioned at 5 μ m and stained for reactivity with rabbit antiserum against mouse Glu6PI- α and indirect immunoperoxidase staining.

To establish a stromal cell line with a dominant selectable marker, the neomycin-resistance gene, encoding a dominant selectable marker that confers resistance to mammalian cells against the antibiotic G418 (G418^r), was transfected on a retroviral vector, first to NIH/3T3 cells and then to cell line GBL/6. Transfection was by calcium phosphate precipitation (Graham et al., 1973, Virol. 52:456). 10 μ g of the vector pZIP-Neo(SV) (X) DNA (see below) was used to transfect mouse NIH/3T3 cells. This vector carries the neomycin resistance gene under the control of the SV40 promoter. Clones producing high viral titers were isolated by selecting the transfected cell line with 625 μ g/ml of G418 (Gibco), and cloning individual G418^r colonies. Of the cell lines producing defective virus, one was chosen from 19 tested because it produced consistently high viral titers of 3×10^6 G418^r CFU/ml and used for infection of long-term marrow cultures. This supernatant was named pSVX-neo^r virus. pSVX-neo^r virus was used to infect GBL/6 cells. A subclone, GBLneo^r was selected in 500 μ g/ml G418 and expanded in vitro. GBL/6-neo^r cells had cytochemical properties and extracellular matrix proteins indistinguishable from those of GBL/6 cells.

GBL/6 cells support growth in vitro of hematopoietic stem cells forming colony-forming units of spleen cells (CFU-S) and of granulocytes, erythrocytes, and macrophage/megakaryocytes (CFU-GEMM) in the absence of detectable growth factors, e.g., interleukin 3 (multi-colony stimulating factor), granulocyte macrophage colony-stimulating factor, granulocyte-stimulating factor.

The hematopoietic support capacity of GBL/6 cells or GBL/6-neo^r cells in vitro could not be attributed to the synthesis of detectable quantities of any known hematopoietin with multi-CSF activity.

Total cellular RNA isolated from the GBLneo^r stromal cell line was used to prepare poly(A)⁺ mRNA by a modification of the guanidine hydrochloride extraction method (Maniatis et al., 1982, Molecular Cloning: A Lab. Manual, Cold Spring Harbor Lab., Cold Spring Harbor, NY). Specific message for interleukin 3 (IL-3) (Fung et al., 1984, Nature 307:233), granulocyte/macrophage colony-stimulating factor (GM-CSF) (Gough et al., 1984, Nature 309:763) macrophage colony-stimulating factor (M-CSF) (Kawasaki et al., 1985, Science 230:291), IL-1 (Auron et al., 1984, PNAS 81:7907), and granulocyte colony-stimulating factor (G-CSF) (Isuchiya et al., 1986, PNAS 83:7633) was identified by hybridization with specific cDNA probes ($> 10^8$ cpm/ μ g) as described in Maniatis et al., ibid. mRNA from GBL/6-neo^r cells was analyzed by RNA blot hybridization for poly(A)⁺ mRNAs of known growth factors. GBL/6-neo^r cells had no detectable poly(A)⁺ mRNA for IL-3, GM-CSF, G-CSF, or IL-1, and did have detectable mRNA for M-CSF. GBL/6 cells produced macrophage colony-stimulating factor constitutively. The support capacity of the GBL/6 cell line may be due either to the ability of M-CSF to trigger release of other CSFs from accessory cells in vivo and in vitro or to another growth factor.

Assays for Hematopoietic Cells

Adherent and nonadherent cells were removed from LTBMCS weekly and assayed in vitro for colony forming ability CFU-GEMM with and without G418 (Johnson et al., 1977, Proc. Nat. Aca. Sci. 74:3879; Nakahata et al., 1982, Proc. Nat. Aca. Sci. 79:3843). The biological activity of each lot of G418 was assessed in colony assays on uninfected cultures. All G418 concentrations reported are in actual weight per milliliter. Progenitor cell assays were done in medium containing 10% BSA (bovine serum albumin), 30% FBS (fetal bovine serum), 10^{-2} M 1-mercaptoethanol with methylcellulose as a semisolid base, and supplemented with three conditioned media: WEHI-3BCM (Ihle et al., 1983, J. Immunol. 131:282), L-cell CM as a source of macrophage colony-stimulating factor (M-CSF) (Stanley et al., 1977, J. Biol. Chem. 252:4305) or 10% pokeweed mitogen-stimulated spleen cell CM and 2.0 μ m/ml EPO (Humphries et al., 1979, Blood 53:746).

In vivo CFU-s assays were done as described by Till and McCulloch (1961; Radia. Res. 14:213). For each group, 5-10 mice were irradiated to 1000 rad, 140 rad/min (cesium source), and injected with 5×10^5 cells/mouse. Mice receiving no progenitor cells died within 12 days of irradiation and had $\leq 0.1 \pm 0.3$ endogenous colonies per spleen.

Engraftment of GBL6-neo^r Cells into Mice

The ability of injected stromal cells to home and stably seed into marrow sinuses *in vivo* was first evaluated by *in vivo* immunohistochemical technique. Glu6PI α stromal cells were identified *in situ* two months after transplant in the RHL marrow sinuses of transplanted mice. Two months after transplantation, neither engrafted nor irradiated nonengrafted control mice demonstrated detectable donor-originating Glu6PI- α cells in spleen, liver, lung, or peritoneal washings. As shown in Table 1, 1 month after transplantation, 26.2% of the adherent stromal cells in marrow cultures explanted from transplanted mice were of donor origin. Adherent stromal cell explants were established, and donor-originating stromal cells were identified at day 18 using specific antiserum against Glu6PI- α alloenzyme marker (Charles et al., 1980, Mol. Cell. Biochem. 29, 11), and immunoperoxidase staining (PAP). These cells are illustrated in Fig. 1, upper panel, which shows adherent marrow cells explanted *in vitro* from the RNA of transplanted mice at 1 month after transplantation. Fig. 1, lower panel, shows marrow cells explanted *in vitro* from control irradiated but non-transplanted mice at 1 month. Donor originating cells were identified *in vitro* using specific rabbit antiserum against murine Glu6PI- α and immunoperoxidase staining. The arrow shows a positive staining cell. Table 1 also shows that the highest percentage of donor-originating cells was 82.8% and 62.5% of total adherent cells in marrow explants 2 and 3 months, respectively, after transplantation. Glu6PI- α cells composed 78% of adherent cells in LTBMCS established from RHLs (13-Gy irradiated) of transplanted mice. Adherent stromal cells from day-70 LTBMCS were trypsinized, replated on coverslips, and processed for PAP. The percentage of donor-originating Glu6PI- α cells was <0.01% in the nonadherent cells and in individual CFU-GEMM colonies derived from these LTBMCS. In contrast, nonadherent hematopoietic progenitor cells harvested from these same LTBMCS had no detectable Glu6PI- α cells. No detectable Glu6PI α stromal cells were identified *in situ* or in LTBMCS from control-irradiated nontransplanted mice (Fig. 1, lower, Table 1).

The recovery of donor-originating GBLneo^r cells in cultures explanted from control-irradiated nontransplanted mice and mice transplanted with GBL6neo^r cells were selected for growth in the presence of G418 (500 μ g/ml). Mice were transplanted with 5×10^5 GB1neo^r cells per mouse as described above. Two months after transplantation the total number of cells recovered were as follows: 5.8×10^6 per RHL (13 Gy) and 8.6×10^6 per left hind limb (3 Gy) from control irradiated nontransplanted mice; 6.5×10^6 per RHL (13 Gy) and 6.9×10^6 per left hind limb (3 Gy) from transplanted mice. Adherent stromal cell explants were established with 5×10^6 cells per dish (60 x 10 mm). Some were fed biweekly with G418 (500 μ g/ml). The number of G418-resistant colonies were scored 17 days after cultures were established *in vitro*. Table 2 shows that G418 resistant stromal cell colonies were found in the explanted RHL marrow of transplanted, but not of control-irradiated mice. Values in parentheses represent percent control G418-resistant colonies calculated as the number of G418-resistant stromal cell colonies per 5×10^6 cells by the total number of stromal cell colonies per 5×10^6 cells x 100.

The physiological function *in vitro* of transplanted GBL6 stromal cells was evaluated. At monthly intervals LTBMCS were established individually from each hind limb of GBL6-transplanted or irradiated-nonengrafted control mice. The functional integrity of the adherent stromal cells was quantitated by measuring the longevity of hematopoiesis as cumulative number of total nonadherent cells and multipotential progenitor cells produced over 70 days *in vitro*. The cumulative number of viable nonadherent cells produced per culture of marrow established at 1, 2, and 3 months after transplant from the RHL of mice transplanted with GBL6 cells was higher than that produced by cultures from irradiated, nontransplanted control mice. The cumulative number of multipotential hematopoietic progenitor cells forming mixed CFU-GEMM colonies, using procedures described below, per RHL culture (13 Gy), established at 1, 2, and 3 month from transplanted mice was $30.5 \pm 3.7 \times 10^2$, $45.6 \pm 2.5 \times 10^2$, and $34.7 \pm 4.2 \times 10^2$, respectively, compared with $5.13 \pm 2.2 \times 10^2$, $7.3 \pm 0.9 \times 10^2$, and $6.04 \pm 0.13 \times 10^2$ for control irradiated, nontransplanted mouse marrow cultures ($P < 0.05$).

The effect of donor stromal cell number on recovery of hematopoiesis in irradiated mice was tested. Mice were inoculated with 1×10^5 , 5×10^5 , or 1×10^5 cells. Long-term bone marrow cultures were established 2 months after GBL6 cell transplantation. Viable nonadherent cells produced per flask over 70 days were quantitated, as were CFU-GEMM from each RHL (13 Gy) or LHL (3 Gy) culture. A detectable chimeric stromal cell population was established with a minimum of 1×10^5 stromal cells.

Production of hematopoietic progenitor cells by RHL cultures from transplanted mice reached 48% of the level seen in cultures from nonirradiated mice compared with 5% in marrow cultures from irradiated nontransplanted mice. An X-ray dose of 3 Gy to the left hind limb decreased hematopoietic stem cell production in marrow cultures from irradiated nontransplanted mice to 35% compared with cultures from nonirradiated mice. However, GBL6 cell engraftment did not detectably increase cell production in LTBMCS

from limbs irradiated at this dose.

The efficiency of repopulation of marrow sinuses of the RHL (13 Gy) by endogenous CFU-S and the recovery of peripheral blood counts was next measured in GBL6 cell-transplanted and in irradiated nontransplanted control mice. Groups of C57BL/6J mice were irradiated in 2-Gy increments from 3 Gy to 7 Gy with the RHL receiving between 10 and 12.5 Gy, as shown in Table 3. Each irradiation group had 5-10 mice per group. All mice received an additional 10-12.5 Gy to the RHL. One group was injected with 5×10^5 GB16 stromal cells per mouse. Values in parentheses are from the group that received no cells but were control irradiated. Six weeks after irradiation and transplantation, a subgroup from each group was sacrificed; cells were flushed from each RHL and assayed for CFU-S. Results are expressed as mean \pm SD of three to five mice. An average of 4.4 ± 2.9 endogenous CFU-S colonies was seen on the spleens of irradiated noninjected mice. A subgroup from each irradiation dose group was injected with 5×10^5 GBL6 cells per mouse. Six weeks after irradiation and transplantation, the RHL from each animal in each dose group was assayed for the number of multipotential stem cells forming CFU-S. Results are expressed as mean \pm SD of at least three mice per group. Nonirradiated mice had a white blood cell (WBC) count of $8.8 \pm 1.6 \times 10^6/\text{mm}^3$, platelet (PLT) count of $173.7 \pm 28.7 \times 10^3/\text{mm}^3$ and erythrocyte (RBC) count of $7.8 \pm 0.05 \times 10^6/\text{mm}^3$.

As shown in Table 3, at lower TBI doses of 3 and 5 Gy, the number of CFU-S-forming multipotential stem cells per RHL was similar in GBL6 transplanted and in control irradiated nontransplanted mice. In contrast, sublethally irradiated (7-Gy TBI) mice transplanted with GBL6 cells showed a significantly higher number of stem cells forming CFU-S in the RHL compared with the number recovered from the RHL of irradiated nontransplanted control mice ($P < 0.01$ compared with values from control irradiated nontransplanted mice.)

The kinetics of recovery of peripheral blood counts in mice after TBI doses of 3 or 5 Gy were similar in transplanted and control irradiated nontransplanted mice. In contrast, a significant recovery of peripheral blood white blood cell count ($6.9 \pm 1.0 \times 10^3$) and platelet count ($112.5 \pm 2.5 \times 10^3$ per mm^3) was seen in 7-Gy-irradiated GB16-transplanted mice compared with irradiated nontransplanted controls (white blood cell count: $4 \pm 0.05 \times 10^3$ cells per mm^3 ; platelet count 50 ± 3 platelets per mm^3 ; $P < 0.05$; Table 3).

The data show that previous irradiation damages the stromal cells of the hematopoietic environment and suggest that niches, freed of endogenous hematopoietic stem cells by the higher dose, provide more efficient seeding sites for injected donor cells.

Expression of the TGF- α Gene in Mice

Homing of Hematopoietic Cells to Irradiated Site

A defective retroviral vector containing a TGF- α cDNA was transfected by electroporation into the hematopoiesis-supportive bone-marrow stromal cell line GBL6, which was shown, in Anklesaria et al., above, to be capable of engrafting *in vivo* to a high dose irradiated site in the syngeneic mouse. The engrafted TGF- α -transfected GBL6 cell line (Gp-TGF α) was found to remain stably associated in the irradiated sites for many months and to produce the ligand TGF- α , a growth factor having functional homology to epidermal growth factor (EGF), and to the same receptor.

The ability of an IL-3 dependent hematopoietic progenitor cell line (32DC13) to engraft and proliferate on the Gp-TGF α cell line *in vitro* was shown by transfecting 32DC13 with a eukaryotic expression vector containing both the selectable gene (*E. coli*-gpt) and the gene encoding the EGF-receptor (EGFR). Expression of the EGFR gene resulted in the activation of a mitogenic signal in response to EGF (see Pierce et al., Science, 1983 239:623). Control nontransfected 32DC13 cells, co-cultivated with either the Gp-TGF α cell line or GBL6, did not proliferate or form cobblestone areas. The EGFR-transfected 32DC13 cells (32D-EGFR) proliferated extensively on Gp-TGF α cells, but not on GBL6 cells, for over 7 weeks *in vitro*. In contrast, hematopoietic progenitor cells from LTBMCS proliferated and produced CFU-GEMM progenitor cells more effectively with GBL6 cells than with Gp-TGF α cells. Thus only cells that could form a receptor/ligand interaction were able to engraft and proliferate extensively *in vitro*. These results are shown in Table 4.

The ability of 32D-EGFR to home to *in vivo* engrafted Gp-TGF α stromal cells was tested. Irradiated C57BL/6 mice were transplanted with 5×10^5 Gp-TGF α cells/mouse. Two months after transplantation, mycophenolic acid resistant 32D-EGFR cells (1.0×10^7 cells/mouse) were injected into Gp-TGF α trans-

planted and control irradiated nontransplanted mice. Donor origin, neo^r TGR- α producing colonies (11.5 ± 2.5 CFU-F limb) were detected in marrow explants from irradiated Gp-TGF α transplanted, but not from control irradiated non-transplanted mice. 32D-EGFR $15.2 \pm 8.0 \times 10^5$ cells/limb were recovered from hind limb explants of irradiated Gp-TGF α transplanted mice, compared to $0.55 \pm 0.10 \times 10^5$ 32D-EGFR cells from hind limb explants of irradiated nontransplanted control mice. These data indicate that marrow stromal cells containing a transfected enzyme-encoding gene became established in the irradiation-prepared bone cavity, and expressed the enzyme following engraftment.

Treatment of Factor VIII Deficiency in Humans

Human Factor VIII-C Gene

The human coagulation Factor VIII-C gene from the cDNA clone pSP64-VIII (Toole et al., 1984, *supra*) can be introduced into any suitable mammalian expression vector, e.g., the vector pCVSVL, and expressed in human stromal cells (gene expression in pCVSVL is described in Kaufmann et al., 1982, *Mol. Cell. Biol.* 2:1304; Clark et al., 1984, *Proc. Nat. Aca. Sci.* 81:2541). The DNA sequence of the coding region of the human Factor VIII-C gene is given in Wood et al., 1984, *supra*, the disclosure of which is to be regarded as hereby incorporated by reference.

The Factor VIII-C gene can be cloned into the PstI site of pCVSVL by excising the gene on a SalI fragment and using synthetic oligonucleotide adapters containing SalI cohesive ends, as described in Toole et al., *supra*. The Factor VIII-C gene in pCVSVL-FVIII will be under the control of the adenovirus major late promoter, as shown in Fig. 3. This plasmid also includes the SV40 polyadenylation site (SV40 poly A), a duplicated SV40 origin of replication (SV40 Ori), and a deletion in pBR322 that enhances replication of such plasmids in animal cells (Lusky et al., 1981, *Nature* 293:79). Selectable genes in the vector are those encoding tetracycline resistance (Tet^R) and mouse dihydrofolate reductase (DHFR). Similar Factor-VIII-C mammalian cell expression vectors are described in detail in Kaufman et al. WO87/04187, pub. July 17, 1987, hereby incorporated by reference.

Assay for Factor VIII Activity

Factor VIII-C activity in culture supernatants can be determined by the Kabi Coatest Factor VIII-C method, modified to afford a sensitivity better than 0.05 mU/ml, and by the one-stage activated partial thromboplastin time (APTT) coagulation assay (Lee et al., 1983, *Thromb. Res.* 30:511) using Factor VIII-C-deficient plasma. The Coatest assay is based on the activation of natural Factor X by Factor IXa; Factor IXa is dependant on Factor VIII. The APTT assay is based on the marked increase in the coagulation activity of Factor VIII in the presence of thrombin. For thrombin activation, samples can be pretreated 1 - 10 min. with 0.2 units/ml thrombin at room temperature. Alternatively, Factor VIII can be quantitated by radioimmunoassay, using an antibody specific for Factor VIII, as described in Wood et al., 1984, *supra*.

Transfection of Murine Cells With the Factor VIII-C Gene into Mice

A vector containing the human Factor VIII-C gene and the neo^r marker gene can be used to transfect murine stromal cells (e.g., containing GBL/6 cells) by electroporation, retroviral vector transfer, or any other suitable conventional vector transfer technique.

Subclonal lines of GBL/6 neo^r cells expressing detectable levels of human Factor VIII-C will be selected *in vitro* by persistence of neomycin resistance, and production in serum-free conditioned medium of those subclones of detectable human Factor VIII-C by Western blot analysis or biochemical assay.

Engraftment Into Irradiated Mice

C56BL/6J male mice are used as recipients. The cells of these animals have three "recipient" markers: Y-chromatin, the beta variant of glucose phosphate isomerase (GPIA), and absence of detectable neomycin

resistance activity. Donor origin cells are derived from the GBL6 neo^r VIII-C-1 clonal line, and have four markers of donor origin: absence of detectable Y-chromatin, detectable immunoperoxidase staining with hetero-antiserum to GPIA, biological neomycin resistance *in vitro*, and detectable production of human Factor VIII-C. A dose response curve for engraftment of these cells into irradiated mice is carried out as follows.

Recipient mice receive 300 cGy total body irradiation (TBI) and 1000 cGy irradiation to the isolated right hind limb (RHL), delivered by a ¹³⁷Cs γ cell 40 irradiator. The day after irradiation, animals are injected intravenously by tail vein with a single cell suspension of 1×10^4 , 5×10^4 , 1×10^5 , or 5×10^5 of a high producing GBL6 neo^r VIII-C clonal line (cells from 10 mice pooled per group). The mice are then returned to their cages and monitored weekly for detectable Factor VIII-C in the plasma. Representative mice are sacrificed and plasma removed by cardiac puncture at different timepoints and assayed by Western blot analysis for human Factor VIII-C, which will be distinguishable from mouse Factor VIII-C by its specific reactivity with monoclonal anti-human VIII-C antiserum (available from several public sources).

At the time of sacrifice of the mice of each dose group, nucleated cells are removed from the right and left hind limbs of each animal and used to set up murine continuous bone marrow cultures, as described above. Bone marrow cultures from the right hind limb would be expected to contain a higher percentage of donor origin, neomycin resistant, stromal cells and continuous marrow cultures from this limb would be expected to produce detectable human Factor VIII-C *in vitro*. Timepoints for explant of marrow from engrafted mice are chosen at one week, two months, four months, and six months after stromal cell transplantation as well as at other intermediate timepoints. Plasma levels of human Factor VIII-C are measured and the level of Factor VIII-C produced over time graphically displayed. The optimum number of stromal cells transplanted is determined in relation to the highest level of Factor VIII-C production in one hind limb.

Irradiation Dosage Testing

If human Factor VIII-C is not detectable in the mouse plasma after reconstitution of one hind limb with donor cells after high dose irradiation of that limb, a sequential "niche" preparation protocol can be used to irradiate both hind limbs and prepare a large niche for engraftment. This approach has been used to detectably change the macrocytic anemia of the S1/S1^d strain of mice in order to achieve a greater percentage of total marrow volume replacement by donor stromal cells. This strain contains a genetic defect in the bone marrow microenvironment resulting in a deficiency of mature erythroid precursors and reduced numbers of pluripotential hemopoietic stromal cells.

A dominant fraction of the marrow microenvironment of S1/S1^d mice was replaced by preparing the mice with either 2 Gy TBI and 20 Gy to both hind limbs (BHL, single schedule) or by sequential irradiation transplantation (multiple schedule). Adult recipient S1/S1^d mice received 1-2 Gy TBI and 10.0-20.0 Gy, to the right-hind-limb (RHL) or both hind limbs (BHL) delivered by a linear accelerator as described in Anklesaria et al., 1987, *supra*. 1 Gy TBI and 10 Gy x-ray doses are preferred due to the relative sensitivity of S1/S1^d mice to TBI. Irradiated mice were transplanted with 5×10^5 stromal cells (GBLneo^r) by intravenous injection (single schedule). For sequential boost irradiation-transplantation studies with S1/S1^d mice, 1 Gy TBI and 10 Gy to the RHL was delivered by linear accelerator on day 0. A single cell suspension of the GBLneo^r cell line was injected I.V. 48 hours later. Two months after the first irradiation schedule, the same group of mice received 1 Gy TBI and 10 Gy to the left hind limb (LHL). Another injection of the GBLneo^r cell line was administered 48 hours later (multiple transplant schedule). Control irradiated nontransplanted mice received 2 Gy TBI and 10 Gy to both hind limbs (BHL).

None of the control irradiated nontransplanted mice survived. LTBMcs established from GBLneo^r transplanted mice (2 Gy TBI and 20 Gy BHL) at 5 months showed increased cumulative CFU-GEMM forming progenitors/flask in both right (298.8 ± 32.7) and left (415.8 ± 36.5) hind limb cultures compared to nonirradiated S1/S1^d mice ($p < 0.05$, Fig. 2). At two or four months after irradiation-transplantation LTBMcs were established from 3 mice/group. In separate experiments, two months after mice received the second irradiation-transplantation, the cumulative number of CFU-GEMM forming progenitors obtained per flask was higher in LTBMcs established from engrafted right (136.1 ± 32) and left (78.6 ± 15.4) hind limbs of GBLneo^r transplanted mice compared to those from control non-irradiated S1/S1^d mice ($p < 0.05$, Fig. 2). In Fig. 2, the symbols for cells from control non-irradiated mice are open circles; from GBLneo^r transplanted RHL-20 Gy mice are open boxes; from GBLneo^r transplanted LHL-20 Gy mice are closed triangles; and from GBLneo^r transplanted LHL-10Gy are closed boxes.

Thus, in S1/S1^d mice, high dose irradiation may help create a "niche" in the marrow cavity to support

transplanted stromal cells, and may also eliminate endogenous stromal cells that suppress hemopoiesis.

Quantitation of Cells and Enzyme Level

The number of donor stromal cells engrafted in vivo is quantitated by the colony assay for neomycin resistance stromal cells described above. This assay is carried out by plating explanted nucleated marrow cells to petri dishes in the presence of a concentration of 100 µg/ml G-418 and scoring of ≥ 50 cell colonies at day 7, following plating of the cells at serial ten-fold dilutions to a limiting dilution.

In order to quantitate the level of production of human Factor VIII-C relative to the number of engrafted stromal cells, the number of colony-forming stromal cells engrafted is determined as described above and the levels of human Factor VIII-C detectable in vivo and in vitro following explant determined, also as described above. These experiments demonstrate detectable human Factor VIII-C in vivo in mice with persistent levels for over six months.

The following procedures were designed to demonstrate that hemophilia-A can be corrected in vivo in humans using a stromal cell engraftment protocol similar to that described above for the murine system. The recipient is prepared with a similar high dose irradiation technique.

Long-Term Human Bone Marrow Cultures

Nucleated marrow cells from patients with hemophilia-A who have previously contracted human immunodeficiency virus infection from HIV-contaminated plasma derivatives are explanted after informed consent has been obtained. These marrows are established in human continuous bone marrow culture using a technique for preparation of suitable medium supplemented with hydrocortisone sodium hemisuccinate.

Marrow from patients to receive transplant are aspirated from iliac crests removing approximately 3×10^8 nucleated cells. The bone marrow harvesting technique is as described by Buchner et al., 1984, Blood 64:630, the disclosure of which is to be regarded as hereby incorporated by reference, and Thomas et al., 1970, Blood 36:507, as modified by Jin et al., 1985, Exp. Hematol. 13:879, the disclosure of which is also to be regarded as hereby incorporated by reference.

Briefly, inoperatively obtained bone marrow specimens from hip surgery are purified free of spicules by mincing with scissors in McCoy's 5A medium, and then single-cell suspensions are prepared by drawing the marrow through progressively smaller gauge needles to a 30-gauge needle. Cultures are established by inoculation of 3×10^7 to 4×10^7 nucleated marrow cells to each Corning 25-cc plastic flask in McCoy's 5A medium supplemented as previously described (Greenberg et al., Blood 58:724, 1981), and containing 12.5% heat-inactivated fetal calf serum, 12.5% heat-inactivated horse serum, and 10^{-6} mol/L hydrocortisone. Cultures are incubated at 33°C and left undisturbed for 7 to 14 days; this time is absolutely required for adherence of hematopoietic stromal cell islands that are associated with long-term hematopoiesis. After this initial incubation, all nonadherent cells are removed by aspiration, and a Ficoll-Hypaque density cut is performed to remove red blood cells. All nucleated cells are then returned to the individual culture flasks in fresh medium. Cultures are then fed twice per week for the first four weeks, and, thereafter, once per week by removal at each feeding of all medium and nonadherent cells and replacement of 8.0 mL fresh medium. The human stromal cells will then be transfected as described above for murine cells, with a vector containing the human Factor VIII-C gene by electroporation, retroviral vector transfer, or any other suitable technique.

The number of stromal cells transfected and expressing Factor VIII-C gene are quantitated by either linkage of the Factor VIII-C gene to a neomycin resistance gene and selected for neo^r resistant colonies, or by subcloning lines in culture and assaying their supernatants for detectable levels of Factor VIII, as described above, or by staining the cells for detectable Factor VIII-C production by immunoperoxidase technique using monoclonal antibody to human Factor VIII-C. A vector transfection system will be chosen which gives a maximum frequency of expression of transfected stromal cells with the VIII-C gene. When the vector pZIP-NeoSV(X), carrying the neomycin resistance gene, is transfected into fresh human stromal cells, neomycin resistance is stably expressed in vitro in approximately 10-15% of cells which are normally sensitive to the neomycin analog, G418. Based on this neo^r experiment, at least 10% of human stromal cells would be expected to stably express the Factor VIII-C gene. If lower numbers are detected, cloned permanent immortalized human bone marrow stromal cell lines, KM101, KM102, KM103, KM104, and KM105 are used (Fitzgerald et al., 1988, Int. J. Rad. Oncol. Biol. Phys. 15:1153) to maximize the likelihood

of transfection and detection of the Factor VIII-C gene in vitro. Factor VIII-C production is quantitated in the clonal lines and compared on a per cell basis with that detected in the GBL6 neo^r mouse line. Toxicology and tumorigenicity studies are performed as follows.

Growth of human stromal cell lines in vitro is associated with potential problems, including activation of Epstein-Barr virus (Rothstein et al., 1985, Blood 65:744), contamination of the stromal cells with pathogens during in vitro culture, including mycoplasma, bacterial or fungal infection, and spontaneous transformation to tumor-inducing lines. Microbial contamination of the stromal cultures is monitored by bacteriological, virological and mycoplasma assays of material. The cells are tested for Epstein-Barr virus nuclear antigen as described in Rothstein et al, supra, and screened by electron microscopy to determine whether there are any evidence of Epstein-Barr virus activation. EBV has never been detected in human bone marrow stromal cells prior to 20-25 weeks of continuous in vitro culture. The mechanism of activation of EBV is not known but is detected at low frequency in control human marrow cultures.

To determine whether spontaneous tumorigenicity has evolved following insertion of the Factor VIII-C gene, stromal cell cultures expressing Factor VIII-C are engrafted at 1×10^7 cells IP, IV, or intrathecally, into nude mice. These animals are monitored for six months to determine whether tumors are detected. Positive controls for these experiments in nude mice include human tumor cell lines HL60 and U937 (Greenberger et al., 1978, Cancer Res. 38:3340). Using these criteria, it is possible to detect evidence of tumorigenicity of human marrow stromal cells expressing Factor VIII-C; 10% and possibly as high as 50-60%, of freshly explanted human stromal cells may express the transfected Factor VIII-C gene.

Patient Preparation

Each patient is prepared for autologous engraftment of his own Factor VIII-C transfected stromal cells by irradiation of one iliac crest. A radiation therapy portal of no greater than 10 cm. wide x 10 cm. long is chosen to incorporate the iliac crest and part of the ischium, unilaterally shielding bladder and rectum, or an angled AP-PA portal is chosen to minimize the volume of intestine, bladder, and other normal soft tissues in the field. A single fraction size of 1000 cGy is delivered to this segment of bone marrow by AP-PA or opposed oblique technique. Fractionated radiotherapy may be more suitable and perhaps more easily tolerated by humans to prepare a high dose irradiated niche. Basic human radiation biology parameters suggest that 500 cGy daily for two days, or 300 cGy daily for ten days, should produce an isoeffect equivalent to 1000 cGy in one fraction (see Fletcher, ed., Textbook of Radiotherapy, 1980, 3rd ed., Lee and Febinger, Phila, Chp. 2, pp. 180-219). (1000 cGy delivered to a single iliac crest would be expected to produce no untoward effects, and is commonly used in palliative radiotherapy of a single bone having painful metastases.) The day after single fraction irradiation of the iliac crest, patients receive intravenous infusion of a single cell suspension of autologous marrow stromal cells expressing the human Factor VIII-C gene. The patient's plasma is then be monitored weekly for detectable human Factor VIII-C levels.

Variations in Protocols

It is expected that human Factor VIII-C production by engrafted autologous stromal cells will be detectable if the chosen injected number of 1×10^8 cells is adequate to selectively seed into the high dose irradiated niche. If Factor VIII-C is not detected in the plasma, it will be necessary to determine whether adequate seeding of the irradiated site has been achieved.

Patients are monitored for in vivo seeding by indium oxine labelling of the donor stromal cells prior to engraftment. A gamma camera whole body scan is carried out 24 hours after engraftment to determine whether the cells have preferentially seeded to the high dose irradiated bone. Assays carried out in the murine model have already demonstrated that the high dose irradiated limb preferentially takes up the injected stromal cells and that the indium label is stable for 48 hours. Doses of indium oxine will be used similar to those used in nuclear medicine lymphoscintogram scanning techniques for lymphocyte homing in Hodgkin's disease, and in red cell sequestration studies for patients with myeloid metaplasia and myelofibrosis.

If Factor VIII-C levels are initially detected in the plasma but then decrease rapidly, it is determined whether the cause is cell graft instability or vector expression instability is the cause of this failure. Vector instability can be corrected by standard methods, i.e., replacing or modifying the vector for expression of the Factor VIII-C gene in vitro; any modifications use data derived from work carried out with fresh explanted human marrow or clonal human marrow stromal cell lines. Graft instability is determined in vivo

studies using S^{35} labelled human marrow stromal cells that have been engrafted *in vivo* and watched for 100 days, which is equal to the half-life of S^{35} . Detectable S^{35} in the high dose irradiated bone initially should be followed by persistent detection in that high dose region for 100 days; a decrease in detectable S^{35} by scanning would suggest instability of cells in the graft site. If this were to occur, other methods for preparation of the niche could be used, although it is expected that the graft will be adequately seeded and stable after 1000 cGy.

Failure to detect adequate levels of Factor VIII-C despite adequate engraftment of stromal cells to the high dose irradiated bone and stable vector expression may be associated with an adequate relative number of stromal cells. This problem may be corrected by preparation of several niches similar to that which was required in the $S1/S1^d$ mouse mutant model. According to this methodology, both iliac crests or one iliac crest and one femur are prepared as larger niches using the same high dose irradiation scheme, fractionated irradiation scheme as described above. Single or multiple inoculations of donor stromal cells each transfected and expressing Factor VIII-C from human marrow cultures are then carried out.

Alternate Methods for Introduction of Factor VIII-C Gene into Human Stromal Cells

Viral vectors containing the Factor VIII-C gene can be introduced into human stromal cells either by infection or by transfection. The recombinant viral DNAs can be propagated in the presence of an appropriated helper virus and packaged into infectious virions, or can be propagated in a packaging-defective viral producing strain. Infectious virus can be produced from, e.g., mouse a NIH/3T3 or ψ_2 cell line which has been transfected with recombinant viral DNA. Cells can be grown as described in Cepko et al., 1984, Cell 37:1057, and infections performed in the presence of 8 μ g/ml polybrene (Sigma).

Retroviruses, comprising single-stranded RNA molecules, can be used for viral infections; their RNA's are converted intracellularly into double-stranded proviral DNAs, which are integrated into the host genome after duplication of one of the termini of the viral sequence to generate the long terminal repeats (LTR). Helper-free stocks of recombinant retrovirus are produced in helper cell lines after transfection with a recombinant retroviral vector. The helper cell lines contain an integrated copy of the retroviral genome. The provirus provides all the transacting functions necessary for replication and encapsidation of a recombinant genome, but is defective in the ability to encapsidate its own RNA.

SV40 derived plasmid vectors can replicate extrachromosomally in monkey COS cells (Gluzman, 1981, Cell 23:175). Vectors that can replicate autonomously in a variety of eukaryotic cells (e.g., human, monkey, dog) are described in Pouwels et al., eds., "Cloning Vectors", 1985, Elsevier Sci. Pub., B.V., the disclosure of which is to be regarded as hereby incorporated by reference. Vectors without a replicon for animal cells usually have a replicon for *E. coli*. Such vectors, also described in Pouwels et al., cannot replicate extrachromosomally in animal cells; transformants can be isolated only if they integrate into the host genome and express a selectable marker. A detailed description of animal cell vectors can be found in Rigby, 1983, J. Gen. Virol. 64:255; Elder et al., 1981, Ann. Rev. Genet. 15:295; and Subramani et al., 1983, Anal. Biochem. 135:1.

Selection of stable transformants can be carried out using a number of genetic markers, e.g., the *E. coli* *gpt* gene encoding resistance to mycophenolic acid, the *neo* gene from TN5 encoding resistance to the antibiotic G418, the *dhfr* sequence from murine cells or *E. coli* which changes the phenotype of DHFR⁻ cells into DHFR⁺ cells, and the *tk* gene of herpes simplex virus, which make TK⁻ cells phenotypically TK⁺ cells.

Stable human stromal cell lines can be constructed that efficiently express the gene of interest by placing the gene under the control of a strong promoter, e.g., the early and late promoters of SV40 (p_{SV40E} , p_{SV40L}), the major late promoter of adenovirus (p_{Adml}), the promoter of a vaccinia gene encoding a 7.5 KD protein ($p_{7.5}$), and the promoter of the murine metallothionein gene (p_{MT}).

Another vector which may be used for transfection of the desired gene is the retroviral vector pN2-FAT (Garver et al., 1987, *supra*), which contains the SV40 early promoter directing transcription of the inserted gene; in this case, the gene encoding human α_1 -antitrypsin. The α_1 -antitrypsin gene can be removed and replaced with a desired gene by digestion with the restriction enzymes *HindIII* and *XhoI* (partial), and modification of these ends or the ends of the inserted gene, if necessary, according to conventional techniques.

Retrovirus-mediated gene transfer can be performed by any suitable method, e.g., as described in Wilson et al., 1988, Proc. Nat. Aca. Sci 85:3014 for introducing a gene into adult rat hepatocytes, using a replication-defective retrovirus.

Table 1

Table 1 Identification of donor-originating stromal cells in bone marrow explanted from transplanted mice				
Time, mo	GB1.6 cells injected per mouse	Donor-originating Glu6Pl-a ⁺ cells, %		
		Adherent stromal cell explants* per limb, %		Adherent† stromal cells from right limb LBMCS
		Right	Left	
1	0	<0.01	<0.01	Not tested
	1 × 10 ⁶	26.2 ± 3.2	28.4 ± 4.4	Not tested
2	0	<0.01	<0.01	<0.01
	1 × 10 ⁵	75.5 ± 9.5	65.0 ± 2	76.7 ± 15.7
	5 × 10 ⁵	82.5 ± 0.5	32.5 ± 3.5	78.0 ± 0
	1 × 10 ⁶	59.0 ± 14.4	16 ± 0	Not tested
3	0	<0.01	<0.01	<0.01
	1 × 10 ⁶	62.5 ± 12.5	<0.01	Not tested

Table 2

Table 2 Recovery of donor-originating GB1neo ^r cells in cultures explanted from transplanted mice				
Group	Stromal colonies per hind limb*, no.		G418-resistant stromal colonies per hind limb†, no.	
	Right	Left	Right	Left
Control-irradiated	65.5 ± 4.1	110.1 ± 10.3	0	0
	78.0 ± 13.0	98.7 ± 13.3	30 ± 2.0 (39 ± 3%)	10.3 ± 2.0 (11 ± 2%)

Table 3

Table 3 Hematopoietic recovery in C57BL/6J mice transplanted with GB1.6 stromal cell line				
Total* body dose, Gy	CFU-S per right hind limb,† no.	Peripheral blood analysis on day 42 after irradiation‡		
		WBC, × 10 ³ /mm ³	PLT, × 10 ³ /mm ³	RBC, × 10 ⁶ /mm ³
3.0	92.0 ± 24.5	8.3 ± 1.5	156.5 ± 6.5	7.7 ± 0.03
	(104.6 ± 26.0)	(7.2 ± 0.3)	(126.5 ± 5.5)	(8.2 ± 0.02)
5.2	108.3 ± 9.2	6.8 ± 0.3	131.5 ± 10.5	8.2 ± 0.05
	(132.5 ± 24.4)	(5.7 ± 0.6)	(108.0 ± 1.0)	(7.4 ± 0.12)
7.0	101.8 ± 11.7§	6.9 ± 1.0¶	112.5 ± 2.5¶	7.5 ± 0.5
	(49.1 ± 9.5)	(4.0 ± 0.05)	(50.0 ± 3.0)	(5.9 ± 1.3)

Table 4

Stromal Cell Lines	Cumulative (7 weeks)			
	Progenitor Cells	"Cobblestone Island"/Flask	Non-adherent Cells/Flask	Progenitor/Flask $\times 10^3$
Gp-TGF α GB1-6	32D-EGFR	1789.3 \pm 106.7	6.17 $\times 10^6$	168.8 \pm 12.0
	32DC13	0	0.06 $\times 10^6$	0
	LTRMCs-d36	N.T.	1.67 $\times 10^6$	2.0 \pm 0.05
	32D-EGFR	0	1.7 $\times 10^6$	32.5 \pm 7.5
	32DC13	0	0.007 $\times 10^6$	0
	LTBMCs-d36	N.T.	8.1 $\times 10^6$	12.8 \pm 0.48

Claims

1. Bone marrow stromal cells which have been transfected with a gene encoding a biologically active enzyme, said cells being capable of adhering to a bone cavity surface of a human patient.

2. Bone marrow stromal cells according to Claim 1, further characterised in that said enzyme is a lymphokine, a growth factor a hematopoietic factor, or a deficient protein normally found in blood plasma, such as a blood coagulation factor.

3. Bone marrow stromal cells according to any preceding claim, further characterised in that said enzyme is human Factor VIII-C.

4. Bone marrow stromal cells according to any preceding claim, further characterised in that said stromal cells are autologous cells.

5. Bone marrow stromal cells according to any preceding claim, and for use in therapy to cause production and secretion into the bloodstream of a human patient of said enzyme, being an enzyme for which said patient suffers a deficiency, by introduction of said cells into said patient.

6. Bone marrow stromal cells according to any of Claims 1 to 4, and a source of irradiation for use in combination in therapy to cause production and secretion into the blood stream of a human patient of said enzyme, being an enzyme for which said patient suffers a deficiency, by introduction of said cells into said patient and irradiation of said bone, preferably a long bone, at a dosage which, in the region of irradiation, is effective to kill hematopoietic cells but not said stromal cells.

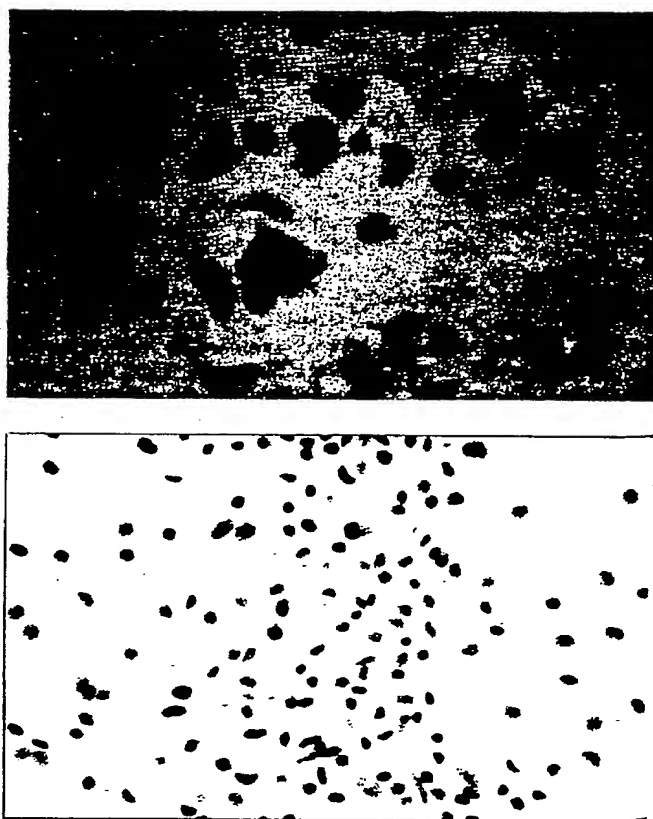


FIG. 1

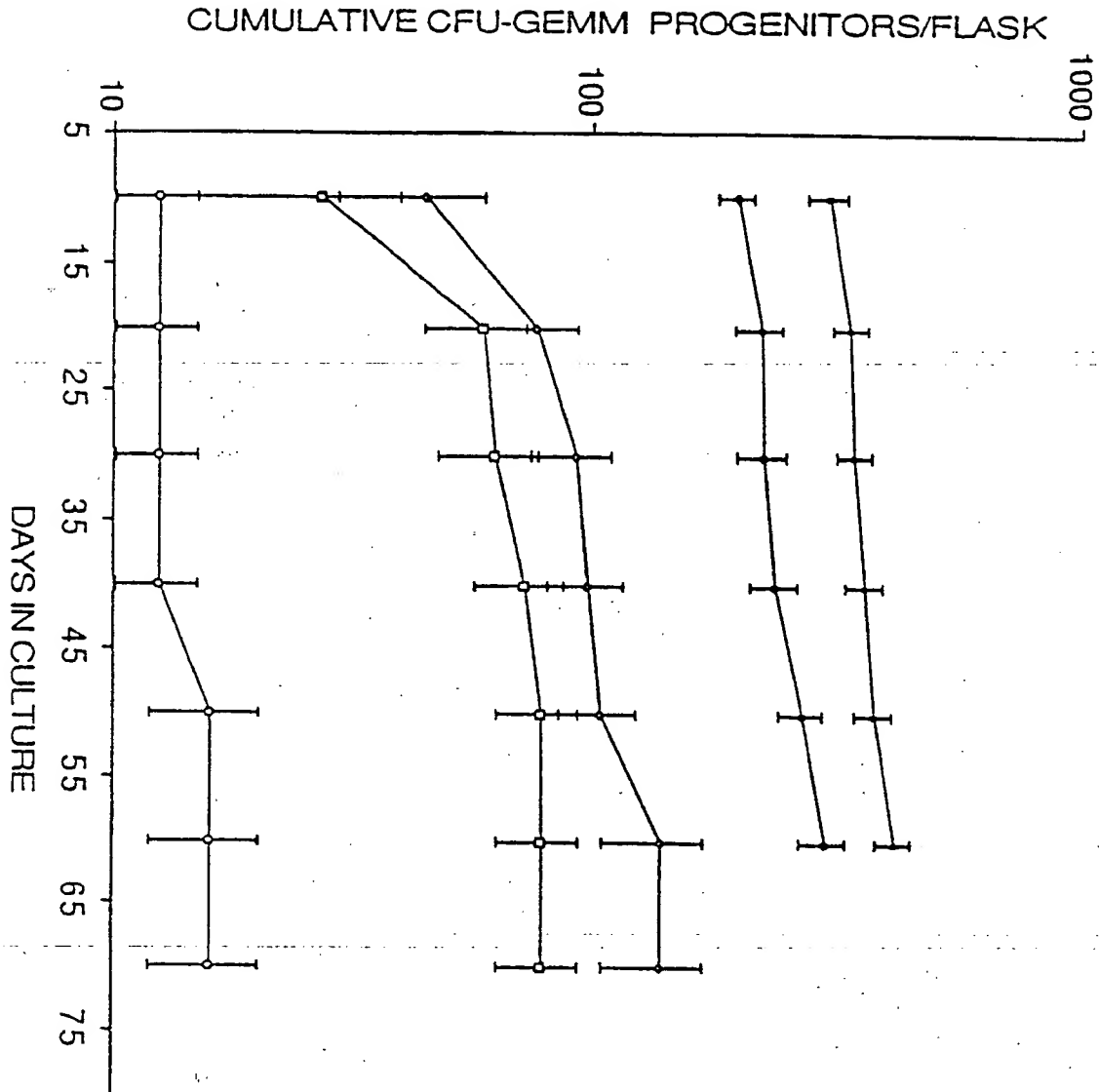


FIG.2

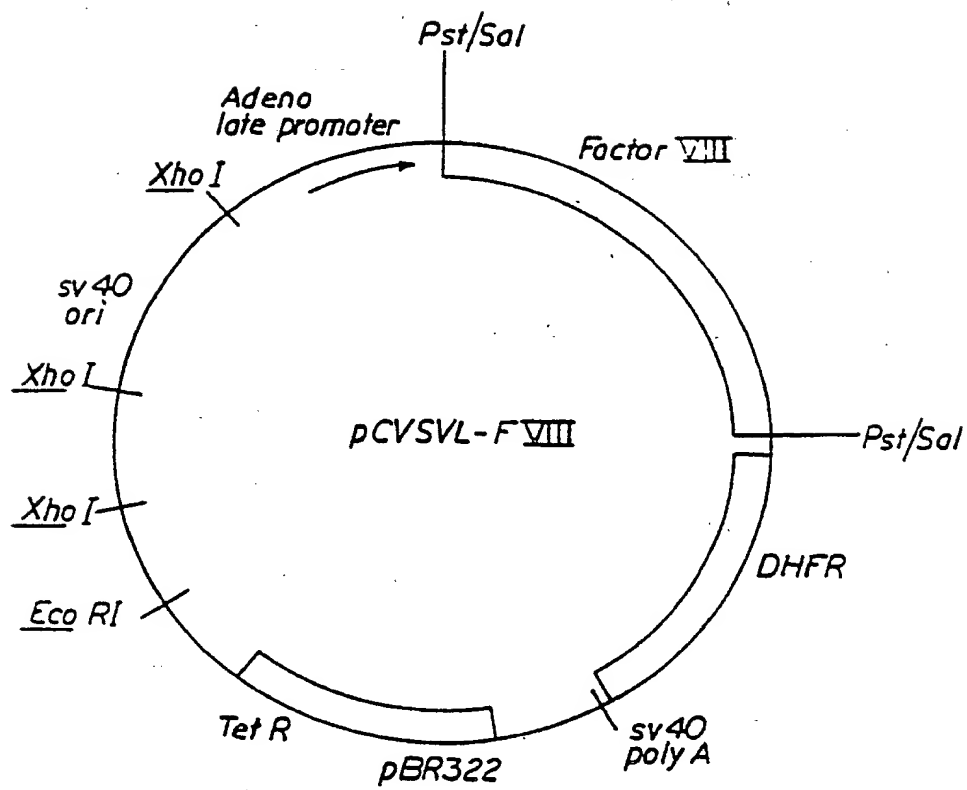


FIG.3



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(11) Publication number:

0 381 490 A3

(12)

EUROPEAN PATENT APPLICATION

(21) Application number: 90301044.5

(51) Int. Cl.⁵: **A61K 48/00**

(22) Date of filing: 01.02.90

(30) Priority: 02.02.89 US 305856

(43) Date of publication of application:
08.08.90 Bulletin 90/32

(94) Designated Contracting States:
DE ES FR GB IT NL

(98) Date of deferred publication of the search report:
15.05.91 Bulletin 91/20

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(54) Gene therapy using stromal cells.

(57) Production and secretion into the bloodstream of a human patient of a biologically active enzyme for which the human patient suffers a deficiency is achieved by introducing into the human patient donor bone marrow stromal cells which have been transfected with a gene encoding the enzyme, so that the introduced cells can adhere to a bone cavity surface of the patient and produce and secrete the active enzyme.

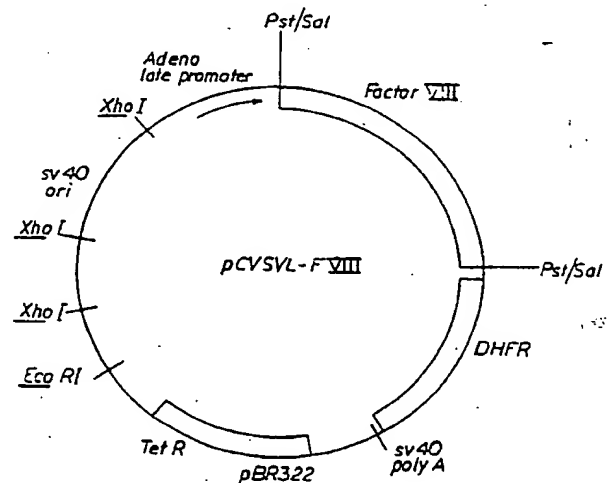


FIG.3



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DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl.5)
Y	INT. J. RADIATION ONCOLOGY BIOL. PHYS. vol. 15, 1988, USA pages 1153 - 1159; FITZGERALD T.J. et al: "Radiosensitivity of permanent human bone marrow stromal cell lines: effect of dose rate." * page 1159 *	1-2, 4-6	A61K48/00
A	* the whole document *	3	
Y	SCIENCE vol. 232, no. 4756, 13 June 1986, USA pages 1373 - 1378; PARKMAN R.: "The application of bone marrow transplantation to the treatment of genetic diseases." * page 1375 *	1-2, 4-6	
A	* the whole document *	3	
Y	THE AMERICAN JOURNAL OF MEDICINE vol. 83, no. 2, August 1987, USA pages 291 - 297; CLINE M.J.: "Gene therapy: current status" * pages 295 - 296 *	1-2, 4-6	
A	* the whole document *	3	TECHNICAL FIELDS SEARCHED (Int. Cl.5)
X,P	ACTA HAEMAT. no. 82, 1989, pages 136 - 143; Holland C. et al: "Infection of hematopoietic and stromal cell in human continuous bone marrow cultures by a retroviral vector containing the neomycin resistance gene." * the whole document *	1-2, 4-6	A61K
A,D	WO-A-8704187 (GENETICS INSTITUTE) * the whole document *	3	
The present search report has been drawn up for all claims			
Place of search BERLIN		Date of completion of the search 13 MARCH 1991	Examiner AVEDIKIAN P.F.
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